

# Chromosome Morphology & Number

**Karyotype** describes the chromosome complement of an individual or species in terms of number, size, and morphology of its chromosomes

**Idiogram** is a diagrammatic representation of the chromosomes, based on observations and measurements in a number of cells

Karyotypes can be based on **mitotic or meiotic** chromosomes and are enhanced by chromosome **banding techniques**. The **largest pair** of chromosomes is usually designated as **number 1** and the rest are numbered in order of their relative diminishing size, to the smallest pair.

# Chromosome landmarks

Primary constriction	The position of centromere defines chromosome arms (telocentric, acrocentric, subterminal, submedian, median, metacentric)
Secondary constriction	Satellites which define the locations of ribosomal RNA genes
Tertiary constriction	Only in some species and often correspond to the sites of cold-sensitive heterochromatic regions

## Mitotic karyotype analysis

Number of chromosomes

Morphology of chromosomes

Heterochromatin vs. Euchromatin

Chromosome Banding

Labeling (i.e. *in situ* hybridization)

## Meiotic karyotype analysis

Number of chromosomes

Morphology of chromosomes (more detailed than mitotic analysis)

Heterochromatin vs. Euchromatin

(more detailed than mitotic analysis)

Chromosome Banding

Labeling (i.e. *in situ* hybridization)

Chromosome pairing behavior

Gene expression (in certain situations)



## Chemicals used to observe chromosomes

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Category	Chemical
Stimulation of cell division	Phytohemagglutinin stimulates lymphocytes to divide
Cell synchronization	Hydroxyurea; Amethopterin (folic acid analog); Cold treatment
Spindle dispersion and spreading of chromosomes	Colchicine, $\alpha$ -bromonaphthalene; 8-hydroxyquinoline; hypotonic treatment (20-30% isotonic)
Staining chromosomes	Feulgen (DNA-specific staining); Giemsa, Carmine

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# Chromosome Banding Techniques

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Name	Basis	Organisms Studied
Q-banding	Banding of AT-specific fluorochromes to DNA	Reptiles, birds, mammals
G-banding	Giemsa staining after incubation in warm SSC or trypsin	Fish, amphibia, reptiles, birds, mammals
R-banding	Giemsa staining after incubation in hot buffer	Mammals
C-banding	Giemsa staining after alkali treatment	Most plants and animals
Replication banding	Incorporation of BrdU during either early or late S phase followed by Giemsa staining	Plants, vertebrates

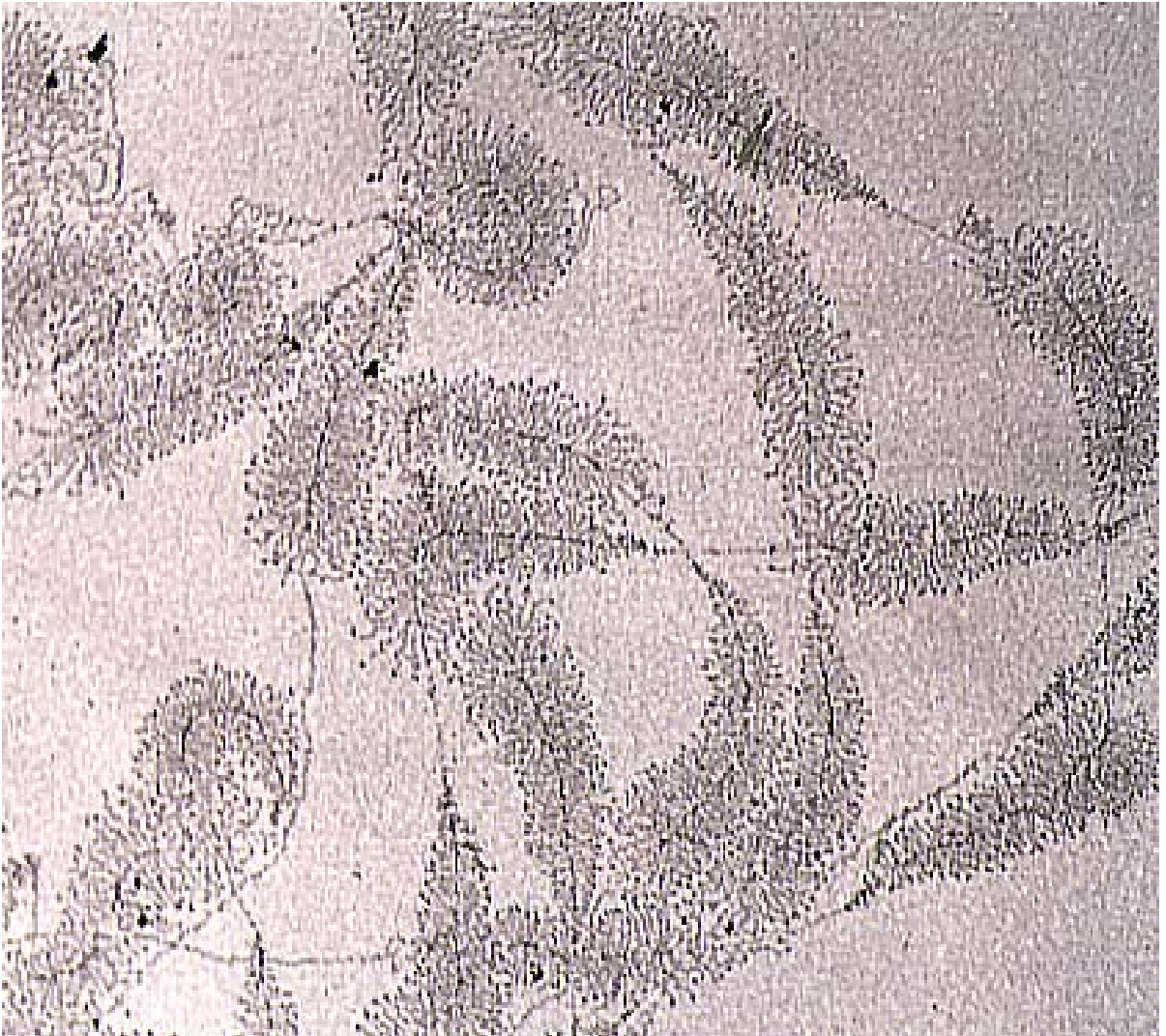
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# Chromosome Banding Techniques

- C-Banding**                      Giemsa stains the satellite-rich centromeric heterochromatin. Therefore C-bands reveal the location of **constitutive heterochromatin**
- R-Banding**                      R bands apparently contain **GC rich DNA** and are the **interbands in G-banded** chromosomes.
- G-Banding**                      Dark bands (G-bands) alternating with light bands are produced in prophase or metaphase chromosomes. The G-bands are region of the chromosome where the DNA is presumably free to stack with the dyes in the stain.
- Q-Banding**                      **AT rich DNA** reacts strongly with quinacrine and enhances its fluorescence in UV while GC rich DNA quenches the fluorescence.

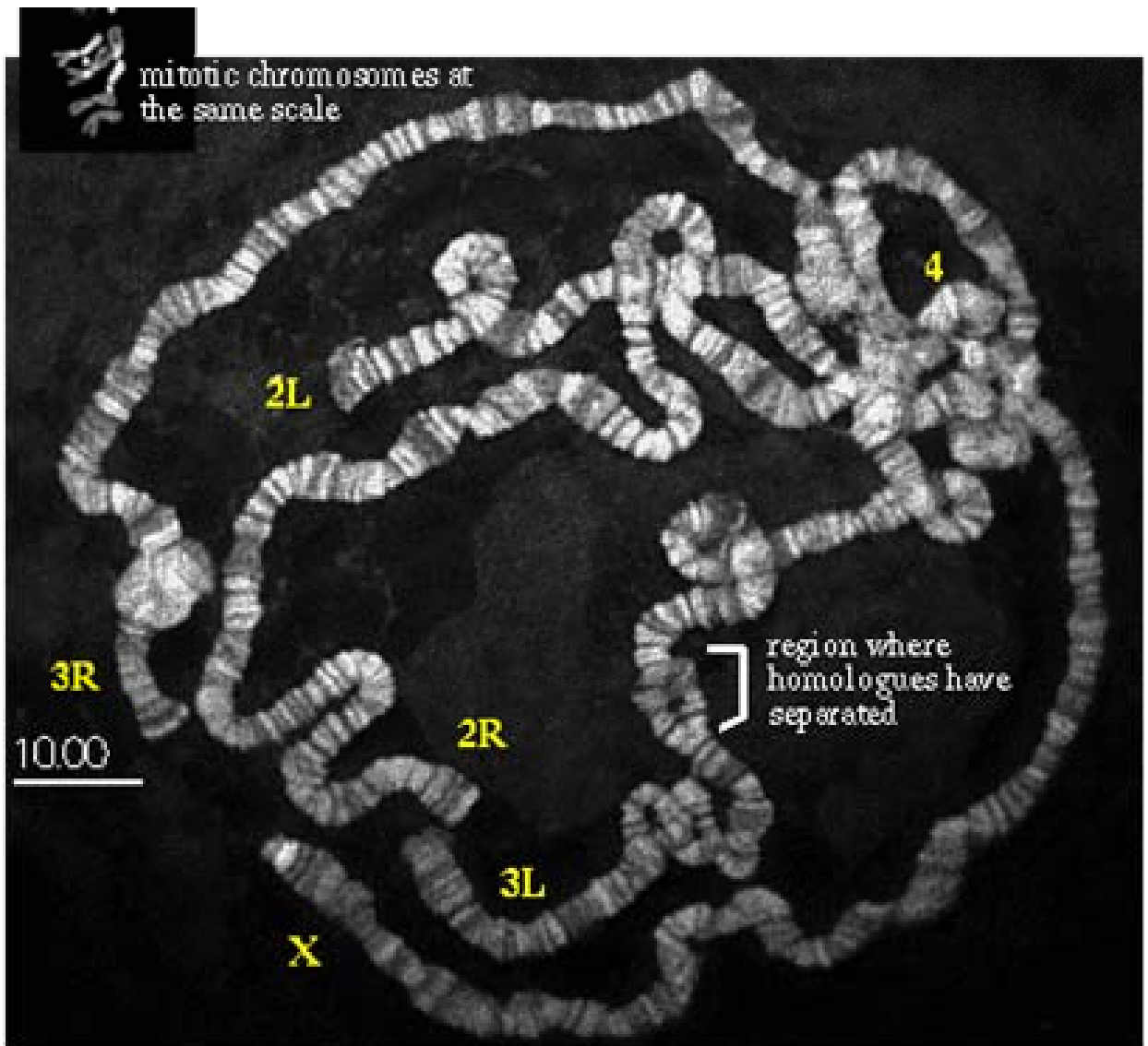
## Lampbrush chromosomes

In the arrested diplotene (dictyoene) chromosomes expand and transcription begins. In oocytes of certain vertebrates this transcription is evident under light microscopy.



## Polytene chromosomes

DNA replication takes place but mitosis is arrested at the  $G_2$  stage. The replicated chromatids of each chromosome remain attached to one centromere. This process of endoreduplication can lead to a final levels of 1024 DNA strands in the polytene chromosomes of *Dorsophila melanogaster* salivary glands.



# Telomere

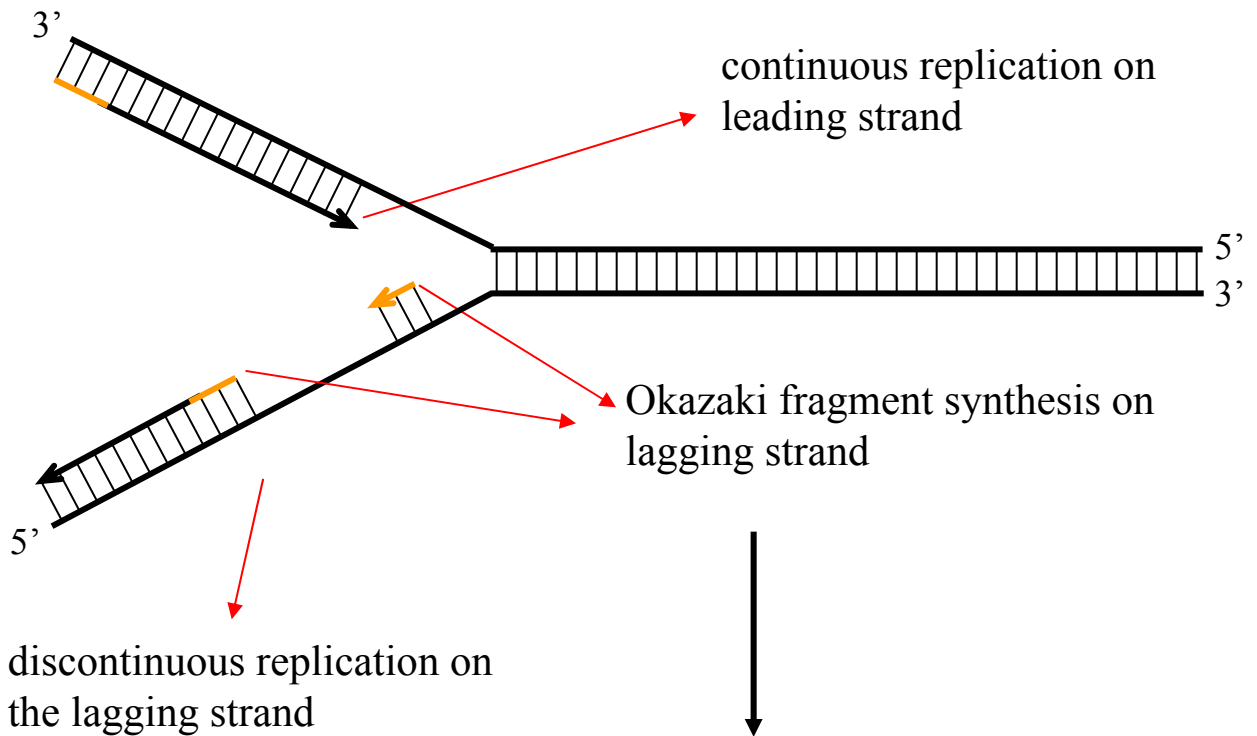
## Why

- ✓ Complete DNA replication.
- ✓ Broken chromosomes are unstable. Telomeres protect chromosomes from end to end fusion.

## Features

- ✓ The DNA strand running from 5' to 3' from the center toward the end of the chromosomes has more G residues than its complement. The G's are clustered.
- ✓ Telomeres are tandem arrays of (T/A)<sub>1-4</sub> (G)<sub>1-8</sub>. For example (TTAGGG)<sub>n</sub> in human.
- ✓ The amount of telomeric DNA varies within and between species.

# DNA synthesis is semi-discontinuous

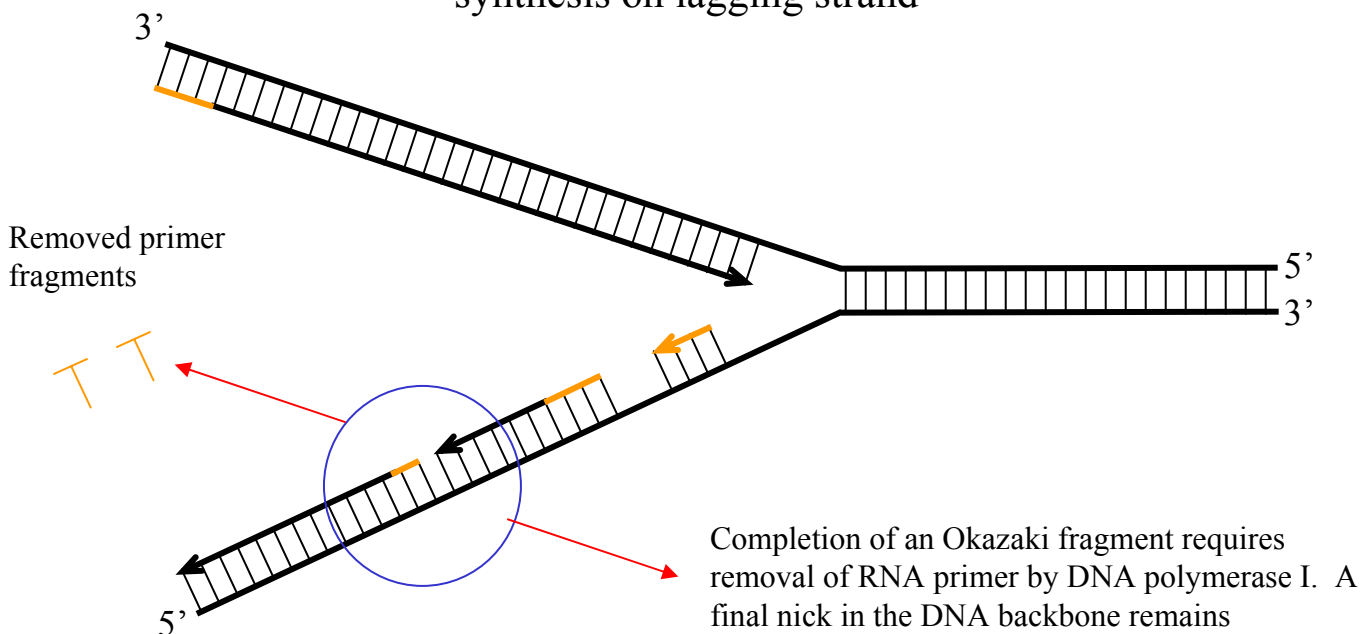


**Continuous replication on leading strand**

+

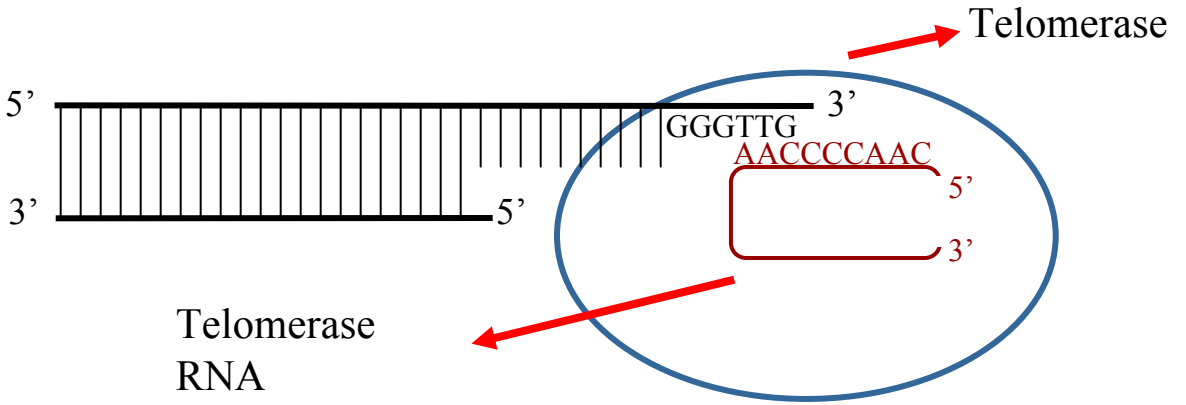
**Discontinuous replication on lagging strand**

RNA primer formation and elongation by DNA polymerase III result in Okazaki fragment synthesis on lagging strand

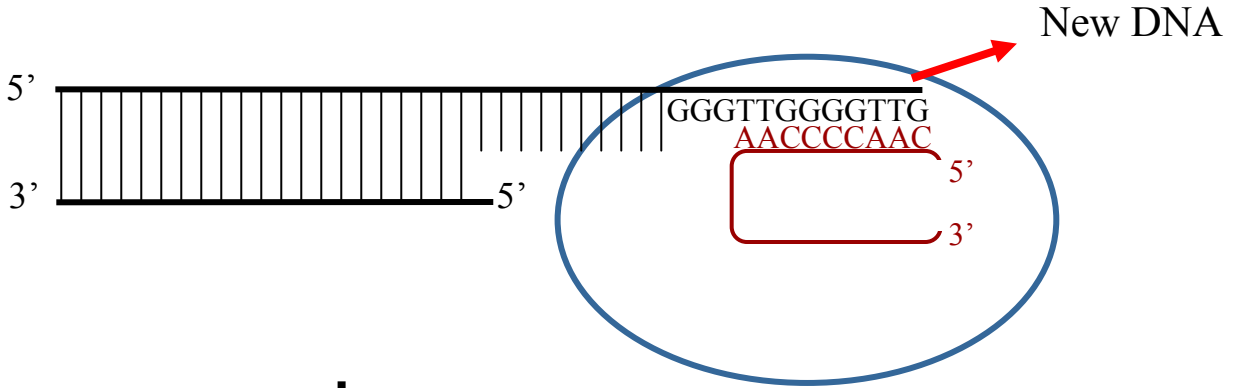


## DNA synthesis is semi-discontinuous

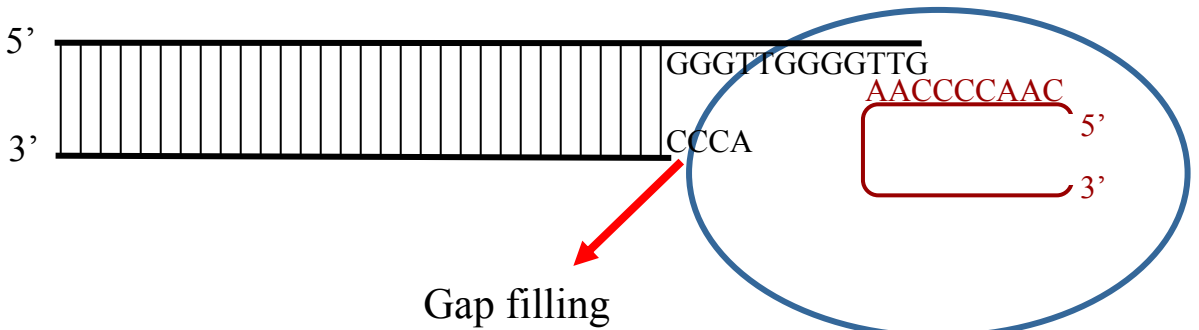
- ✓ DNA polymerase replicates DNA in 5'-3' direction using 8-12 base stretch of RNA to prime DNA synthesis
- ✓ One strand of a linear chromosome will be replicated to the very end, the other will have a 8-12 base gap generated by the removal of the primer.
- ✓ Telomeres act as substrate for replication of the ends and prevent exonucleolytic degradation of chromosomes.



Reverse transcription  
(telomere extension)



Translocation of telomere  
and gap filling by DNA  
polymerase I

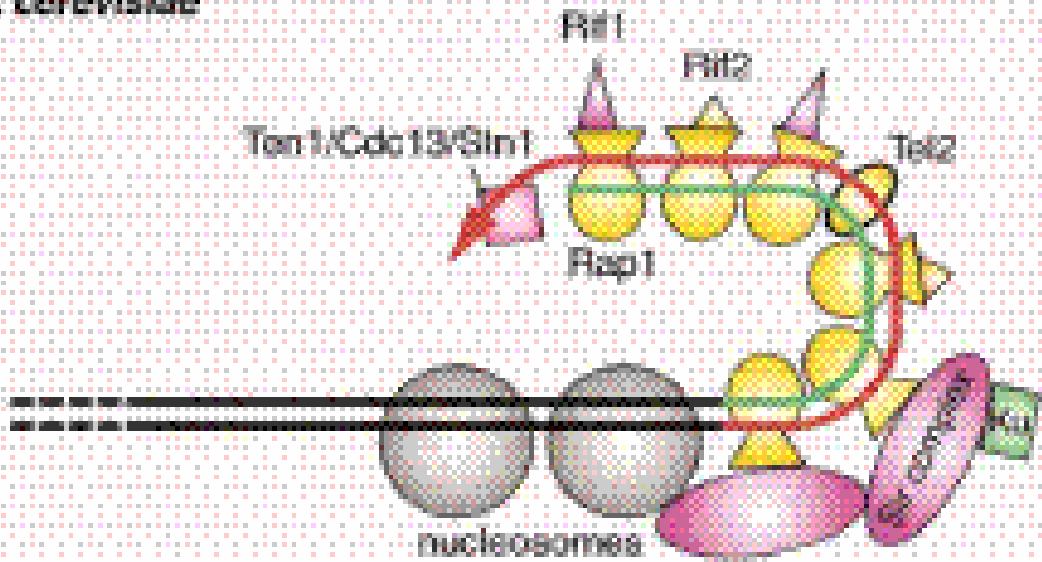


# Telomerase

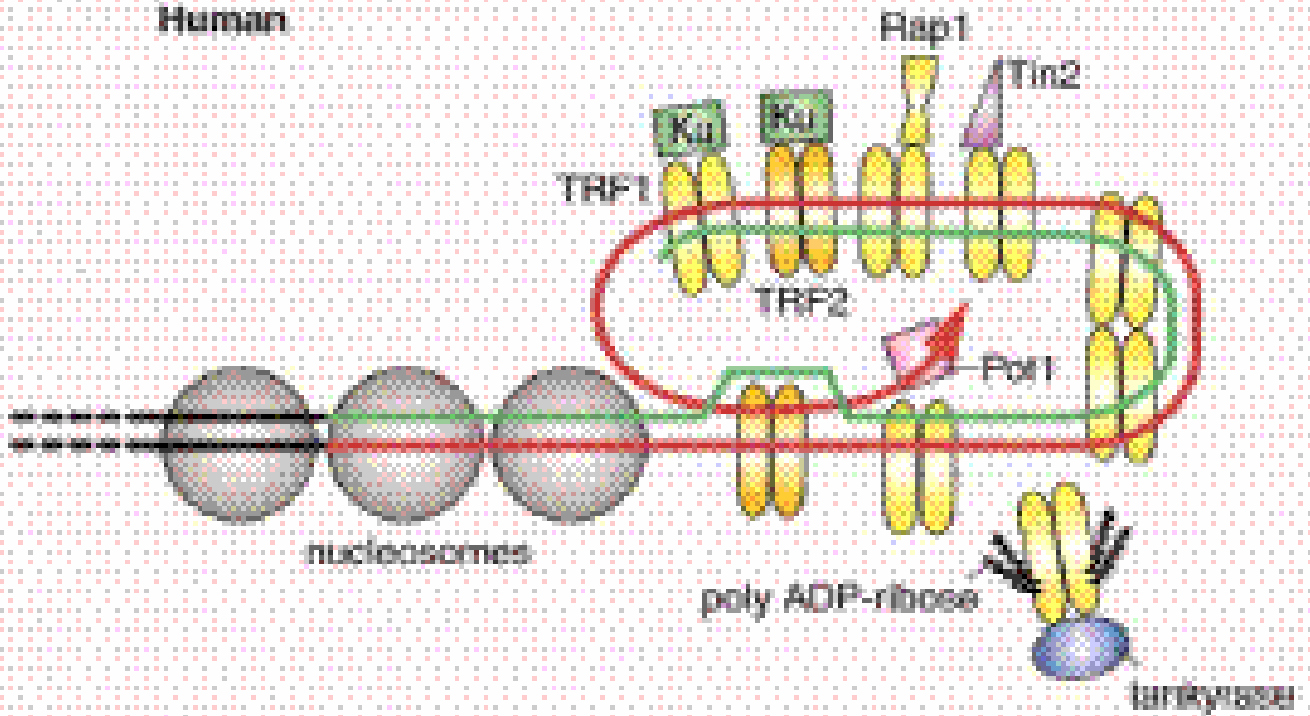
- ✓ Specialized reverse transcriptase
- ✓ Ribonucleoprotein with activity depending on both RNA and protein components.
- ✓ Uses RNA, not DNA to template synthesis of the telomeric DNA
- ✓ The expected product of telomerase mediated replication is a duplex molecule with a single strand G tail
- ✓ Telomerase RNA determines the sequence of telomeric DNA
- ✓ Telomerase independent mechanisms exist for telomere maintenance in humans (no telomerase activity found in somatic cells).



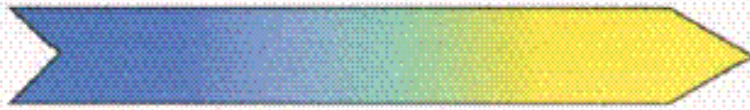
*S. cerevisiae*



Human

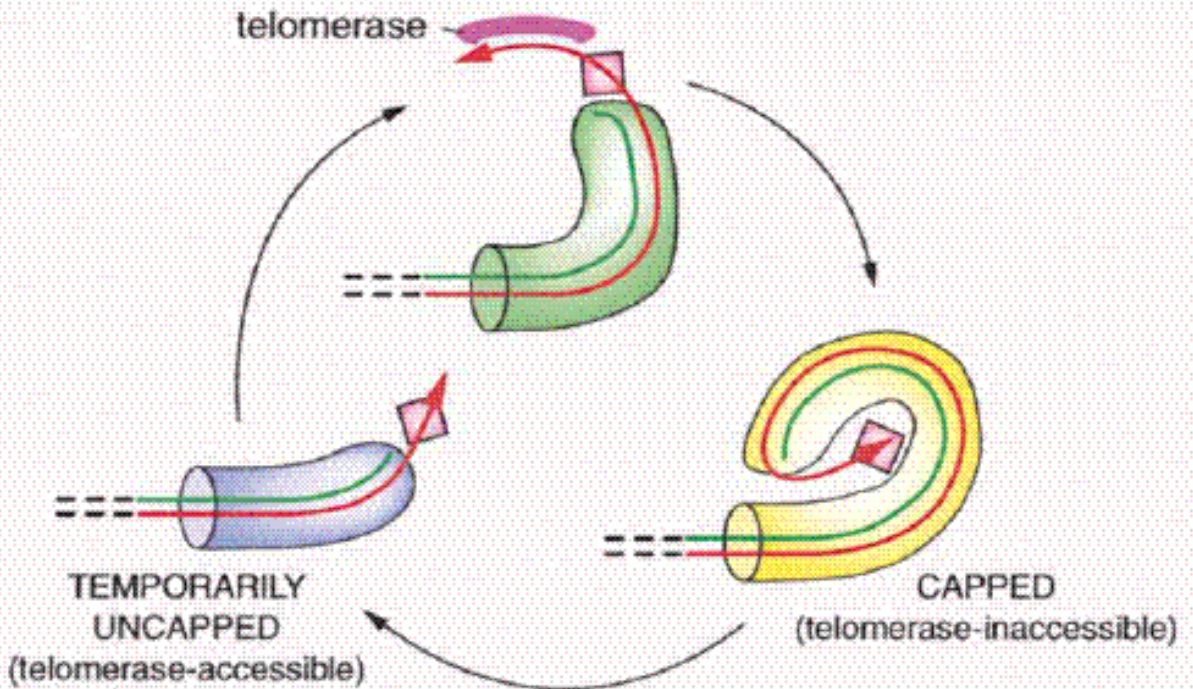


Low telomerase activity; high amounts of telomerase-binding proteins

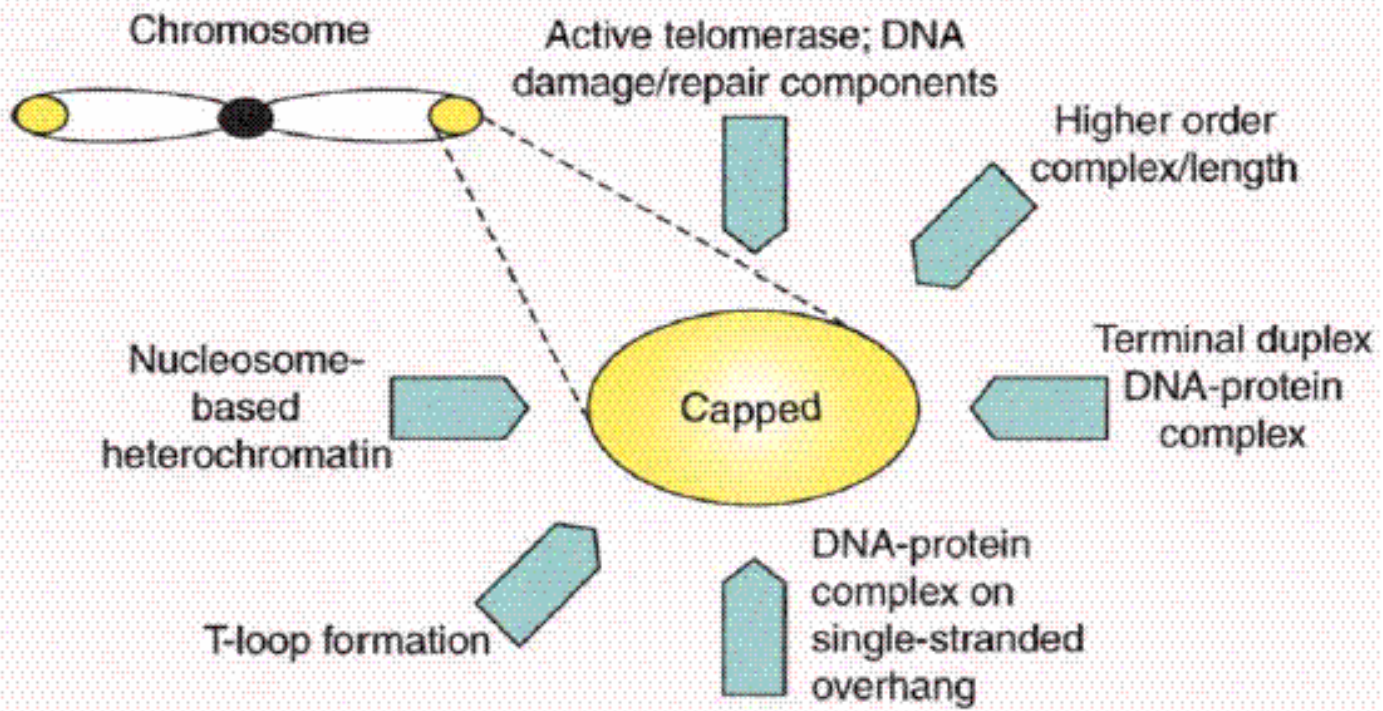


High telomerase activity; mutations in telomere-protective proteins or their telomeric DNA binding sites

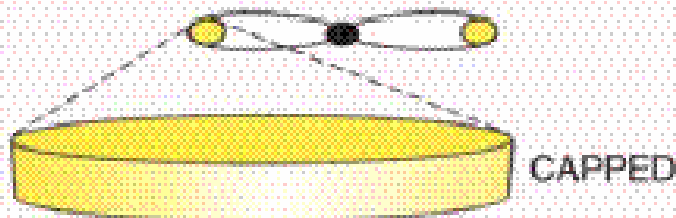
Lengthen telomere by telomerase



Shorten telomere by incomplete DNA replication, nuclease action



Chromosome

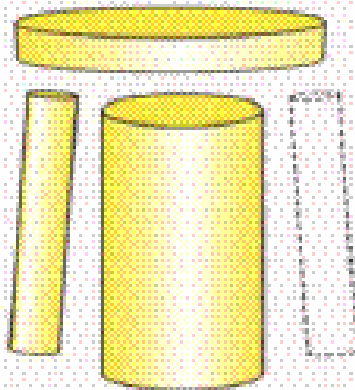


Terminal repeat complexes;  
T-loop formation

Active telomerase;  
DNA damage/  
repair components

Higher order  
complex (length)

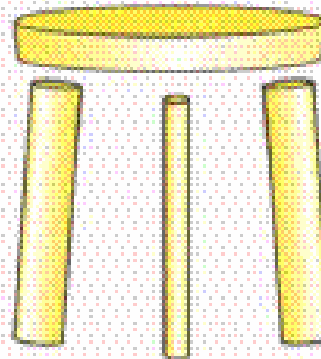
CAPPED



No active  
telomerase

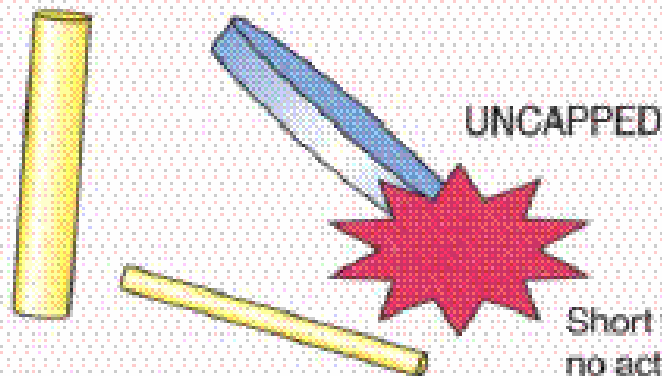
Long telomere

CAPPED



Active  
telomerase

Short telomere



Short telomere,  
no active telomerase

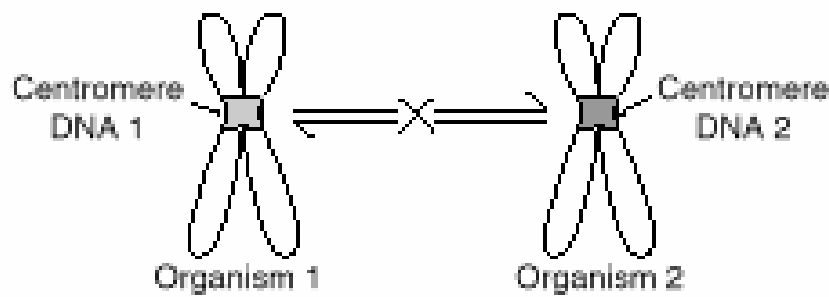
# Centromere

- ✓ CEN locus- DNA sequence that specifies the centromere location
- ✓ Site of formation of the kinetochore
- ✓ Final locus of sister chromatid pairing in mitosis and meiosis
- ✓ Cell cycle checkpoint control
- ✓ DNA contains repetitive sequences
- ✓ Centromeric proteins have been identified which are associated with DNA in the centromere region (CENP-A, B, C and D).

Observation

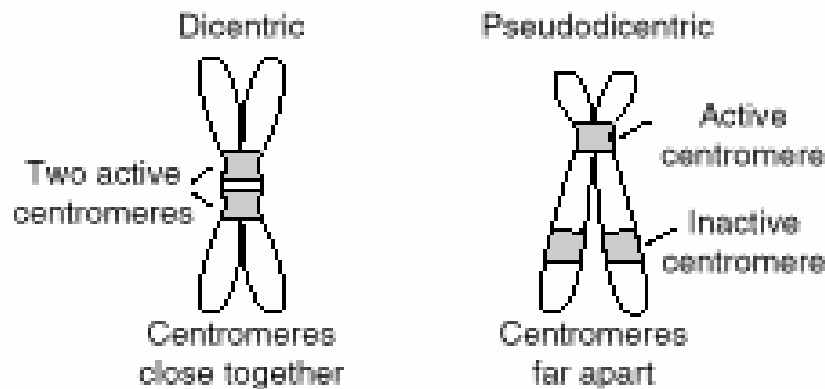
Mechanistic implication

(a) Centromere sequences are not conserved



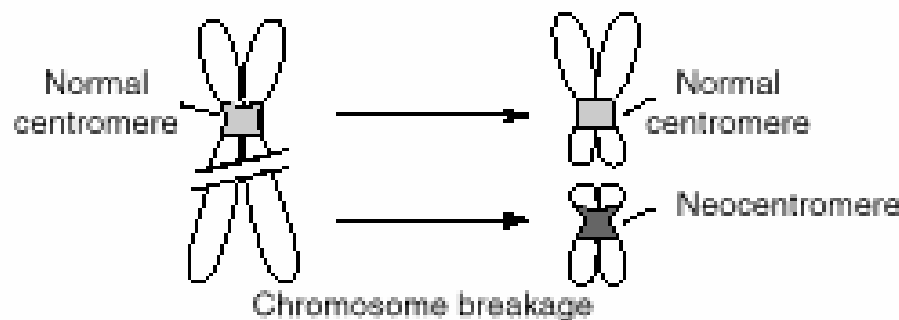
Recognition of widely different centromeric repeat sequences

(b) Dicentric/pseudodicentrics with one or two active centromeres



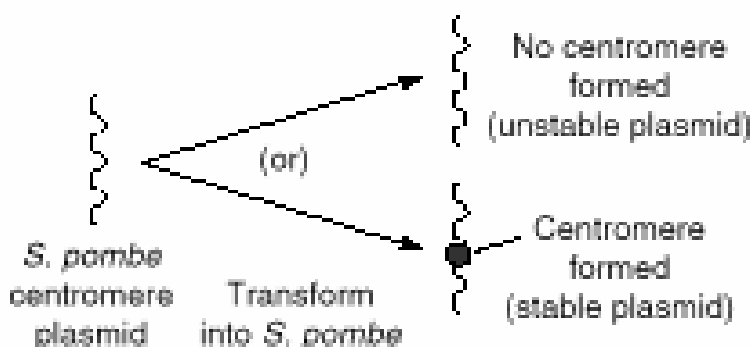
Normal centromeric DNA may not form kinetochore  
Epigenetic influence is distance sensitive

(c) Neocentromeres can form from non-centromeric DNA



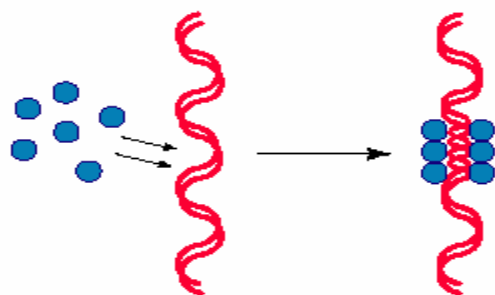
Recognition of a variety of genomic DNA sequences

(d) Variable centromere competence on the same DNA

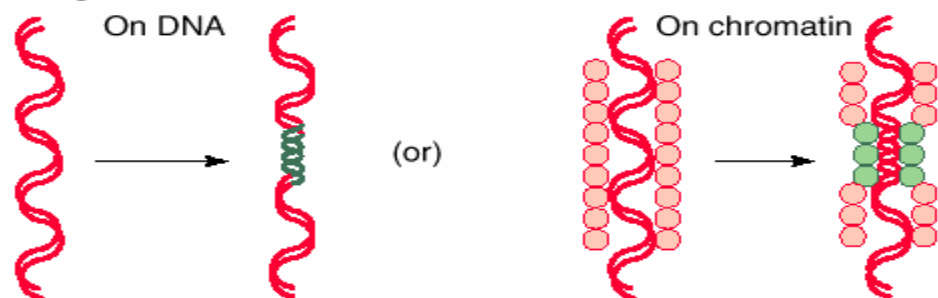


Sequence-independent centromerization

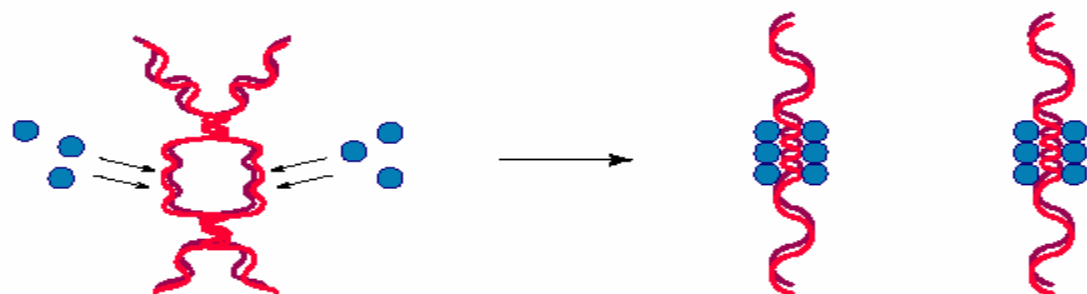
**(a) Protein marking**



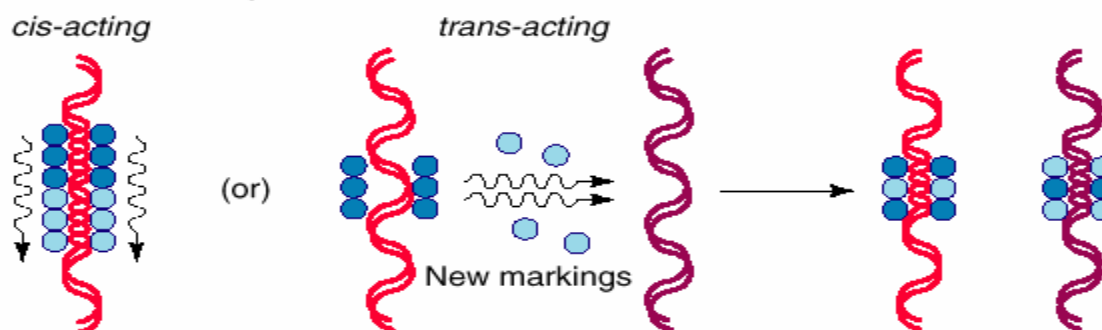
**(b) Chemical marking**



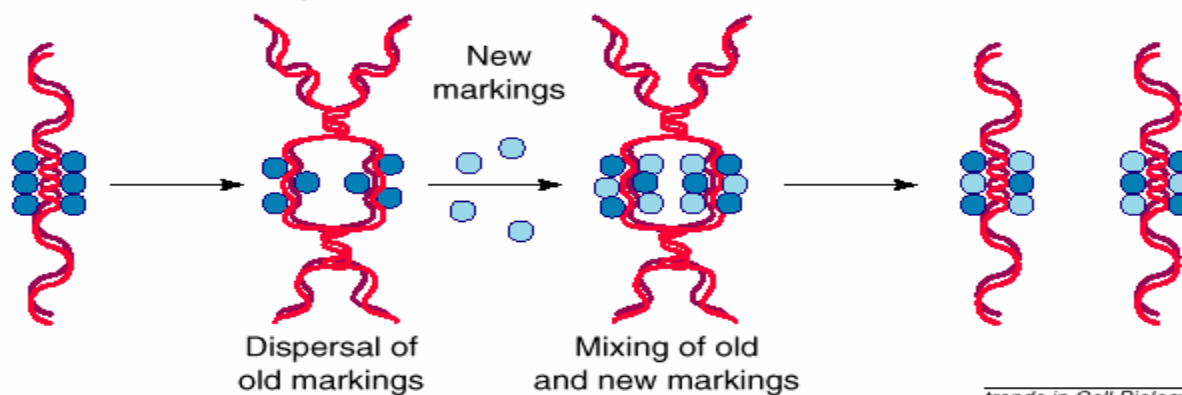
**(c) Synchronization of centromere replication and protein marking**

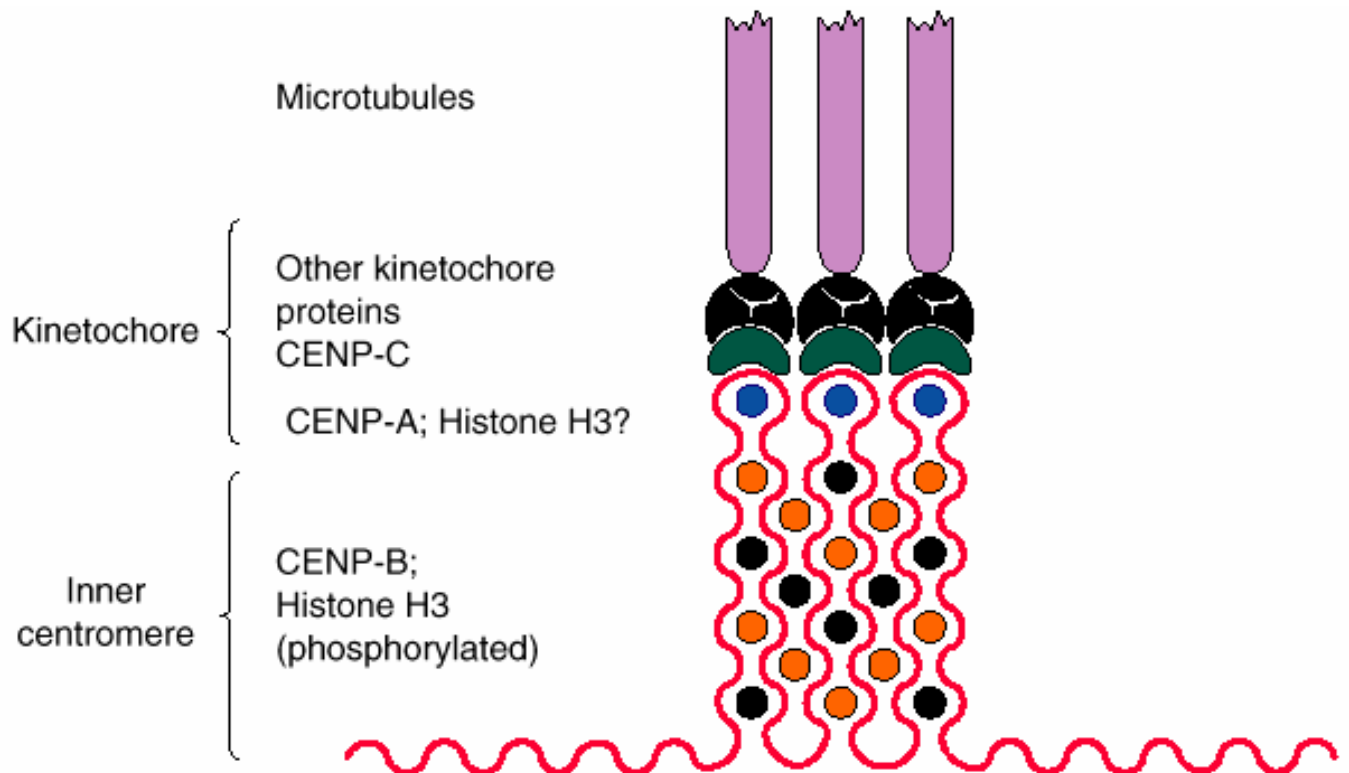


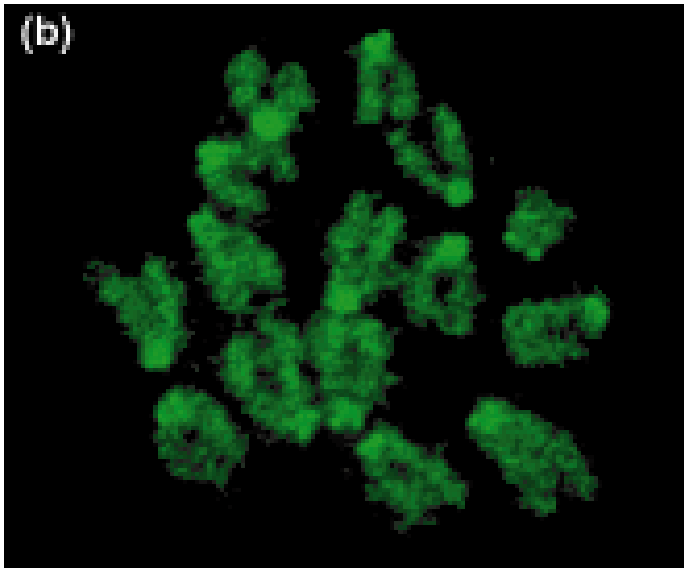
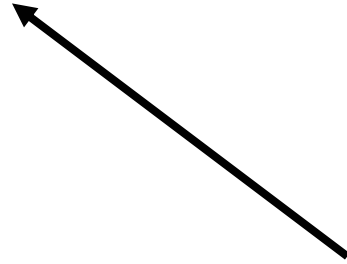
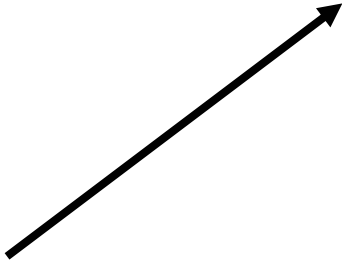
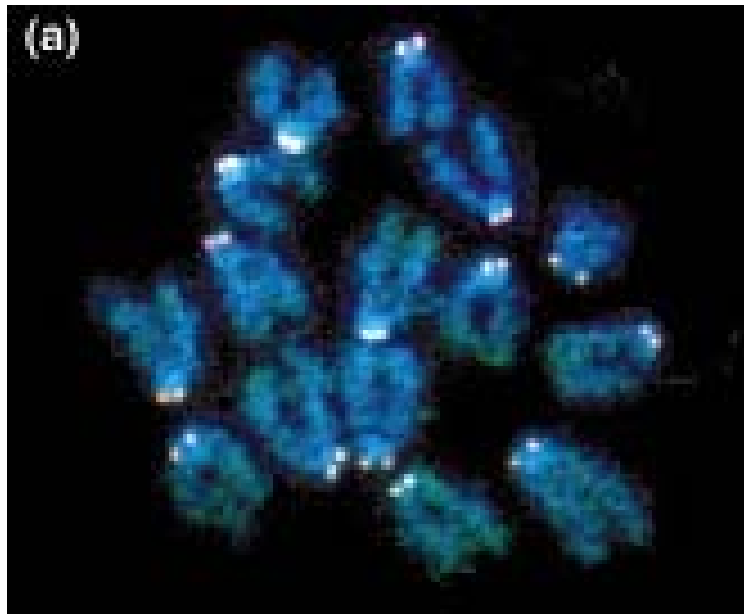
**(d) Chromatin templating**



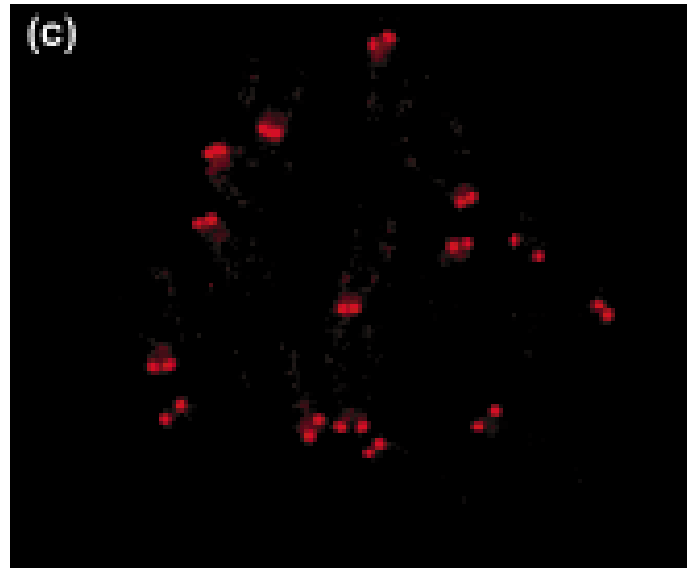
**(e) Centromere propagation**







**Poly (ADP) polymerase (PARP)**



**CENP-A / CENP-B**